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Nutritional state of the pollen beetle parasitoid *Tersilochus heterocerus* foraging in the field

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Abstract Many laboratory studies have demonstrated that parasitoids of various species depend on sugar sources such as nectar or honeydew. However, studies about nectar acquisition by parasitoids foraging in the field are scarce. *Tersilochus heterocerus* Thomson is one of the more abundant and widespread parasitoids of the pollen beetle (*Meligethes aeneus* F.) but nothing is known about the nutritional ecology of this species. In this study we examined the nutritional

state of *T. heterocerus* at the time of emergence and at various time periods throughout the season while foraging in the field using high-performance anion-exchange chromatography. We found that: (i) *T. heterocerus* emerge with relatively small amounts of sugar, composed mainly of trehalose, glucose and fructose; (ii) the first parasitoids caught just after they appeared in the field at the beginning of oilseed rape flowering had already consumed significant amounts of sugar reserves; and (iii) the total amount of sugar at the end of flowering was always significantly higher

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than the total amount of sugar at the beginning of flowering. This study provides valuable insights into the acquisition of sugar in the field by the parasitoid *T. heterocerus* and suggests that nectar acquisition takes place in the oilseed rape field or in the surrounding landscape.

Keywords Oilseed rape · *Tersilochus heterocerus* · Parasitoid · Nectar · Carbohydrate · HPAEC · Conservation biological control

Introduction

Parasitoid wasps are important biological control agents in agroecosystems (Hawkins et al. 1997). Various studies have reported that floral resources can provide key energetic elements to parasitoids which can lead to enhanced pest biocontrol (Berndt et al. 2002; Tylanakis et al. 2004). It has been demonstrated that some parasitoids are particularly dependent on carbohydrates as energy sources and that the availability of suitable sugar sources can play a major role in adult parasitoid longevity, fecundity, searching behavior and population dynamics (Lavandero et al. 2005; Winkler et al. 2006; Lee and Heimpel 2008). The major carbohydrate sources for adult parasitoids are believed to be sugar-rich materials derived, directly or indirectly, from plants, such as nectar and honeydew (Jervis et al. 1993; Wäckers and van Rijn 2005). For host-feeding parasitoids, host haemolymph can also provide sugar but is generally considered a poor source because levels of carbohydrates are generally very low (Steppuhn and Wäckers 2004).

Sugar feeding enables parasitoids to maintain a high level of glycogen and slows down the rate of lipid decline which enhance egg production, female fecundity, longevity and foraging ability (Lee and Heimpel 2008). It has been shown that carbohydrate levels increase with sugar consumption and are thus considered a good indicator of the parasitoid's nutritional state in the field (Casas et al. 2003; Steppuhn and Wäckers 2004).

Complex landscapes with a high proportion of semi-natural habitats have been found to support higher natural pest control by parasitoids than simple landscapes with small amounts of semi-natural habitats (Bianchi et al. 2006). This positive effect of landscape complexity on biological control is assumed

to be due to the provision of key resources for natural enemies such as food, overwintering sites or alternative hosts (Landis et al. 2000). A thorough understanding of the biology and patterns of resource utilization by parasitoids is necessary to interpret parasitism and population dynamics at the landscape level (Costamagna et al. 2004). Such knowledge is, however, usually lacking for numerous parasitoid species. In this paper, we focus on food resource acquisition by *Tersilochus heterocerus* Thomson (Hymenoptera: Ichneumonidae) foraging in the field.

Tersilochus heterocerus is one of the three most abundant and widespread parasitoids of the pollen beetle [*Meligethes aeneus* F., (Coleoptera: Nitidulidae)], a major oilseed rape (*Brassica napus* L.) pest, and can cause a high level of mortality in the pest population (Ulber et al. 2010). It is a univoltine, koinobiont, larval endoparasitoid of pollen beetle larvae (Jourdheuil 1960). Adults appear in the oilseed rape crop at the beginning of flowering (i.e. end of April, beginning of May in Western Europe) and females lay their eggs into host larvae located in buds or open flowers. Then, pollen beetle larvae drop to the ground and the parasitoid completes its larval development in a few days, killing its host and pupating in the top centimetres of the soil. *Tersilochus heterocerus* overwinters as diapausing adults in a cocoon and adults emerge the following spring from the soil of the previous-year oilseed rape fields (Ulber et al. 2010). According to Jourdheuil (1960), this species does not feed during imaginal activity and sexual maturation occurs during diapause and depends solely on lipid reserves accumulated during larval development within the host body. Females emerge with a high quantity of already mature eggs and can therefore rapidly parasitize pollen beetle larvae in the field (Jourdheuil 1960).

Despite the observations by Jourdheuil (1960), various authors think that adult parasitoids of the pollen beetle probably search for nectar in the field and in the surrounding environment, though no proof has yet been found (Thies et al. 2003; Jönsson et al. 2005; Zaller et al. 2009; Rusch et al. 2011). One of the potential explanations of the positive effect of landscape complexity on parasitism rates of the pollen beetle is that parasitoid populations may benefit from the higher nectar availability found in complex landscapes, suggesting nectar exploitation by parasitoids during the adult stage (Thies et al. 2003; Zaller

et al. 2009; Rusch et al. 2011). Although there is evidence that adults of various parasitoid species generally use sugar sources to satisfy their energetic needs (Jervis et al. 1993; Steppuhn and Wäckers 2004), nothing is known about the nutritional state of pollen beetle parasitoids, despite the fact that they are major biological control agents receiving increasing attention (Ulber et al. 2010).

In the context of conservation biological control there is now a need to examine the nutritional state of pollen beetle parasitoids during foraging in the field. Moreover, collecting data about the nutritional state of biological control agents could contribute to a better understanding of patterns of predation or parasitism at the landscape level. The aim of this study is to provide detailed information on the feeding history, the nutritional state and the type of food consumed by *T. heterocerus* foraging in the field.

Materials and methods

Field sites

The experiment was conducted in 2009 and 2010 in four agricultural fields (two different fields for each year) (field area; 2009: 4.7 and 3.1 ha; 2010: 5.7 and 16.5 ha) located in the same region in north-western France in a sub-Oceanic climate zone (49°05'N, 1°15'E). The landscape of this region is relatively complex and mainly characterized by a mosaic of arable land and semi-natural habitats such as small forest fragments, hedgerows and grasslands. The mean proportion of semi-natural habitats in a 2,000 m radius circular sector around each field was $36.14 \pm 19.3\%$ and was mainly composed of forest and grassland [calculated using ArcGis software (Version 9.2, ESRI)]. Oilseed rape (cv. 'Kadore') fields at least 4 km apart were sown by the participating farmers between the 28 August and the 3 September for both years.

Insect sampling

Tersilochus heterocerus foraging in the field were collected by sweep net sampling during flowering of oilseed rape, between 11:00 and 16:00 on two different dates for each year. Parasitoids were sampled in an area not treated with pesticides and located in the

centre of each field. All individuals were sampled on the same day for each collection date. The two different dates were selected according to plant development (Fig 1) and climatic conditions (sunny, non windy and dry days). Because *T. heterocerus* appear in the crop at the beginning of flowering and are visible until the end of flowering (Ulber et al. 2010), we sampled adults in the field at these two stages. We sampled adult parasitoids at the beginning of the flowering between growth stages GS 60 and GS 62 (BBCH). The end of flowering was defined as the period between GS 65 and GS 67. The respective sampling dates each year were: 24 April (four days after the beginning of flowering) and 5 May in 2009 (16 days after the beginning of flowering) and in 2010, 29 April (eight days after the beginning of flowering) and 17 May (26 days after the beginning of flowering) (Fig 1). Immediately after sampling, all parasitoids were stored in plastic bags at $-22\text{ }^{\circ}\text{C}$ for subsequent identification, enumeration and analyses.

In both years, sweep net sampling was performed before the first day of flowering to ensure that no parasitoids were already foraging in the fields. Newly emerged adult specimens were obtained from cocoons (containing diapausing adults) collected in the soil of an experimental plot located at the INRA (French National Institute for Agricultural Research) experimental station, Grignon, France. Oilseed rape was grown in the plot in the previous year and was not tilled after harvest in order to maximise *T. heterocerus* survival during the following winter (Nilsson 2010). To determine the initial sugar levels at the time of emergence, *T. heterocerus* within cocoons were

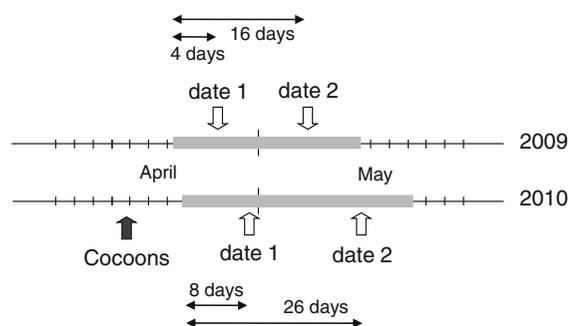


Fig. 1 Sampling dates of *T. heterocerus* at the time of emergence (cocoons, black arrow) and during (date 1 and date 2, white arrows) flowering of oilseed rape (in grey) for 2009 and 2010. Numbers of days indicate the number of days between sampling and the beginning of flowering

sampled 29 March 2010 just before emergence. Cocoons were extracted by washing 20 undisturbed soil cores under water. The cores were taken at equally distributed points across the field. Seventy males and 70 females *T. heterocerous* were extracted by dissecting collected cocoons and were analyzed as unfed individuals. Of these unfed *T. heterocerous*, 25 individuals of each sex were used for preliminary analysis to calibrate the minimum number of individuals required for quantifying sugar content by high-performance anion-exchange chromatography (HPAEC) analysis (data not shown).

HPAEC analysis

All parasitoids were conserved in an Eppendorf tube containing 1 ml 70 % ethanol and stored at room temperature until analysis. The ethanol was evaporated at room temperature and all individuals were then weighed separately with a microbalance before analysis. For each field, we analysed three replicates of pooled individuals of each sex at each date. Preliminary analysis revealed that sugar concentrations contained in one specimen were too low to quantify and using pools of five specimens was the minimum number required to allow quantification (data not shown). When possible (i.e., cocoons specimens and field-caught individuals during 2010) we used more individuals per replicate to increase the detection of the analysis. Thus, for cocoons specimens, we used three replicates of 15 individuals for each sex. For field-caught *T. heterocerous*, we used three replicates of five individuals for each field and sex in 2009 and three replicates of ten individuals for each field and sex in 2010. Pooling individuals to allow sugar quantification in parasitoids is a widely used approach in nutritional ecology studies (Casas et al. 2003; Lee et al. 2006; Winkler et al. 2010). To compare the measured concentrations between treatments, sugar amounts were expressed relative to the parasitoid weight. Parasitoids were placed in 500 μ l of ultrahigh-quality water (18.2 m Ω) and crushed using an automatic homogenizer (IKA[®] RW14 basic). Specimens were then centrifuged at 13,000 r.p.m. for 10 min. We collected 500 μ l of the supernatant and passed it through a 0.2 μ m mixed cellulose ester membrane filter using a 1 ml syringe (Hamilton[®]). For each

sample, 120 μ l was injected into HPAEC Dionex ICS-3000 equipment. Separation of carbohydrates was carried out on a CarboPac PA-1 guard column (4 \times 50 mm) and a CarboPac PA-1 anion-exchange column (4 \times 250 mm) using H₂O (eluent A) and 250 mM NaOH with 4 mM sodium acetate (eluent B) under isocratic mode with the A/B ratio (48:52, v/v) as mobile phase at a constant flow rate of 0.7 ml min⁻¹. Carbohydrates were monitored by pulsed amperometric detection. Curves of sugars were obtained for sorbitol, mannitol, trehalose, glucose, fructose, melibiose, saccharose, melezitose, raffinose and erlose by injecting calibration standards with different concentrations of 2.5, 5, 7.5, 10, 12.5 and 15 nmoles of these sugars. The concentrations of each sugar were analyzed using the program CHROMELEON version 6.7. The limits of detection (LOD) ranged from 2.6 μ g ml⁻¹ for sorbitol to 7.4 μ g ml⁻¹ for erlose. The limit of quantification (LOQ) was calculated according to the limit of detection, following: LOQ = 3 \times LOD.

Physiological parameters

We used two different physiological parameters to describe the nutritional state and the feeding history of field collected parasitoids (Steppuhn and Wäckers 2004; Desouhant et al. 2010). We used the overall sugar content resulting from the sum of all amounts of all sugar types and the ratio between the sum of all amounts of all sugar types except fructose and the sugars sum (defined as the ρ ratio by Desouhant et al. (2010)). Laboratory approaches on several parasitoid species have demonstrated that the overall sugar content provides information on the time since the last meal (parasitoids that fed recently having a higher total amount of sugar than earlier-fed or starved individuals) and that the ρ ratio enables the differentiation between fed and unfed parasitoids (unfed individuals having a higher index than fed ones) (Steppuhn and Wäckers 2004; Desouhant et al. 2010). In the present study we only measured carbohydrate amounts as they are a good indicator of the acquisition of food in non host-feeding parasitoids (Desouhant et al. 2010), like *T. heterocerous*. Because larger individuals were assumed to have greater nutrient stores, the measured sugar concentrations were expressed relative to insect weight.

Statistical analysis

The data were not normally distributed and the variance heterogeneity was high. Therefore, we compared sugar content and ρ ratio between dates for each year and sex using the non-parametric Kruskal–Wallis ANOVA ($\alpha = 0.05$). Post-hoc comparisons of mean ranks between dates were made following Siegel and Castellan (1988). Differences in sugar content and ρ ratio between sexes from the same date were analysed using the Mann–Whitney U test. All statistical analyses were performed using the statistical program R (version 2.8, 2008).

Results

Sugar levels at the time of emergence

The total amount of sugars in *T. heterocerus* parasitoids at the time of emergence was $0.74 \pm 0.25 \mu\text{g}$ (mean \pm SE; $n = 6$) per mg of parasitoid. We found no statistical difference between the total amount of sugars contained in males and females in cocoons specimens (Mann–Whitney U test, $W = 1$; $P = 0.2$). Males contained an average of $0.39 \pm 0.26 \mu\text{g}$ total sugars per mg of parasitoid ($n = 3$) and females contained an average of $1.08 \pm 0.36 \mu\text{g mg}^{-1}$ of parasitoid ($n = 3$). However, in terms of composition, the HPAEC sugar spectrums revealed differences between males and females at the time of emergence. Trehalose was the main sugar found in females. HPAEC sugar spectrums revealed that males contained trehalose, glucose and fructose.

Sugar levels of males and females in the field

The total amounts of sugar in parasitoids sampled in the field are presented in Fig. 2. We found a significant effect of sampling date on sugar content of both males ($\chi^2 = 7.56$, $df = 2$, $P = 0.02$) and females ($\chi^2 = 10.73$, $df = 2$, $P < 0.01$) in 2009. At the beginning of flowering (i.e. 24 April; date 1), the total amount of sugars for both sexes was significantly lower than their respective values in cocoons specimens (Fig. 2) because no sugars were found at quantifiable levels, although trehalose and saccharose were detected during the analysis. At the end of flowering in 2009, the sugar content of males rose to 0.97 ± 0.49

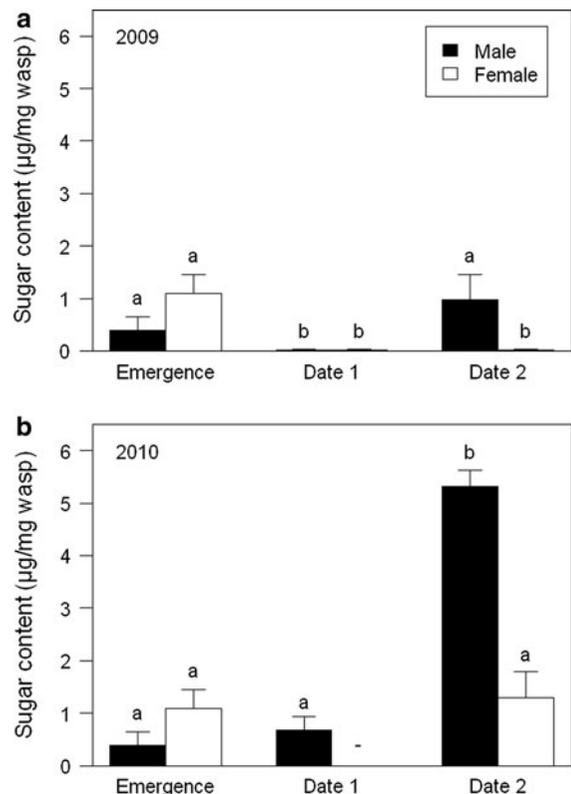


Fig. 2 Mean total sugar amounts (\pm SE) measured in adult *T. heterocerus* at the time of emergence and when foraging in the fields in 2009 (a) and 2010 (b). Sugar content values at the time of emergence are from the same specimens and are reported in both graphs to make comparisons easier. Hyphen in the female column indicates that no specimens were sampled at this date. Different letters indicate significant differences between dates for each sex ($\alpha = 0.05$)

$\mu\text{g mg}^{-1}$ of parasitoid (mainly glucose and fructose) but no sugars were quantifiable for females (Fig. 2). At the end of flowering, the total amount of sugars for males was significantly higher than the sugar content of males at the beginning of flowering but there was no significant difference when compared to the sugar content of males at the time of emergence (Fig. 2).

In 2010, we found a significant effect of sampling date on sugar content of males ($\chi^2 = 10.42$, $df = 2$, $P < 0.01$) and no effect on sugar content of females. Only males were captured at the beginning of flowering and the total amount of sugars at that time was not significantly different from the amount of sugars in the male specimens at the time of emergence (Fig. 2). Sugars in males at emergence were mainly composed of glucose and fructose although trehalose, melibiose

and saccharose were also detected in insufficient amounts for quantification. At the end of flowering during 2010, the total amount of sugars in males was significantly higher than sugar content of males at the time of emergence and at the beginning of flowering when it rose to $5.32 \pm 0.31 \mu\text{g}$ per mg of parasitoid (Fig. 2). Females sampled at the end of flowering in 2010 were found to contain $1.29 \pm 0.50 \mu\text{g}$ of total sugar per mg of parasitoid (Fig. 2). This was significantly lower than the sugar content of males on the same date but not significantly different from the female specimens at the time of emergence or from the sugar content of males at the beginning of flowering. Sugars from males and females at this date were mainly composed of glucose and fructose although melibiose, saccharose and erlose were also detected in insufficient amounts for quantification.

When considering the two years together and normalising sampling dates according to the number of days after the beginning of flowering to account for variability in sampling dates between years, we found that the total amount of sugars in parasitoids increased with the number of days after the beginning of flowering. No sugars were found in parasitoids caught four days after the beginning of flowering (date 1 in 2009), while the total amounts of sugars in males sampled eight days (date 1 in 2010) and 16 days (date 2 in 2009) after the beginning of flowering were 0.68 ± 0.26 and $0.97 \pm 0.49 \mu\text{g mg}^{-1}$ of parasitoid respectively. At eight and 16 days after the beginning

of flowering, the total amounts of sugars in males were not significantly different from those in unfed individuals. After 26 days from flowering (date 2 in 2010), we found significantly higher amounts of sugars than at any other date after the beginning of flowering for both males and females ($\sigma\sigma$: $5.32 \pm 0.31 \mu\text{g mg}^{-1}$; ♀♀ : $1.29 \pm 0.50 \mu\text{g mg}^{-1}$).

The ρ ratio values and patterns of feeding history in the field

The ρ ratio for specimens at the time of emergence was 0.55 ± 0.29 for males and 1.00 ± 0.00 for females (Fig. 3). In 2009, the ρ ratios at the beginning of flowering for males and females and at the end of flowering for females were zero as no sugars were quantifiable. There was a significant effect of sampling date on the ρ ratio values for males ($\chi^2 = 7.2132$, $\text{df} = 2$, $P = 0.02$). At the end of the flowering period the ρ ratio value was 0.50 ± 0.10 for males (Fig. 3). This value was significantly different from the ρ ratio at the beginning of flowering but not significantly different from the ratio of males at the time of emergence.

In 2010, we found no significant differences between the ρ ratios of males at the time of emergence, the beginning of flowering and the end of flowering ($\chi^2 = 1.36$, $\text{df} = 2$, $P = 0.50$) (Fig. 3). The ρ ratio of females at the end of flowering in 2010 was not significantly different than the ρ ratio of females at the time of emergence ($\chi^2 = 2.88$, $\text{df} = 1$, $P = 0.09$).

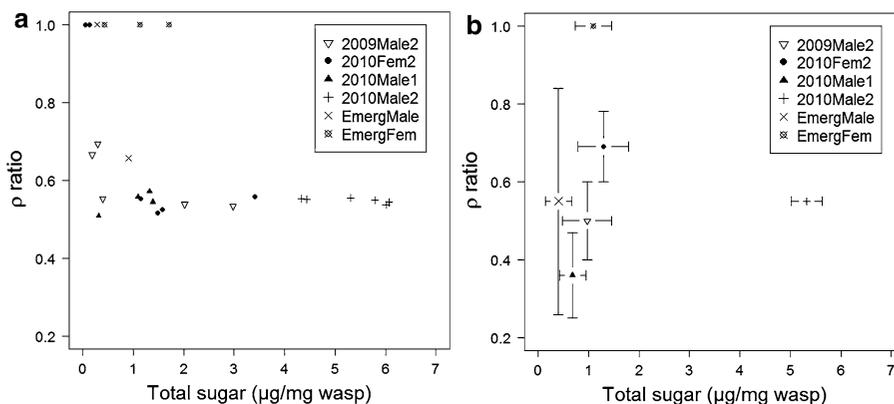


Fig. 3 Plot of the overall sugar content ($\mu\text{g mg}^{-1}$ of wasp) versus the ρ ratio for each pool of individuals sampled at the time of emergence and sampled in the field and for each treatment: sex (male/female), sampling date (date 1/date 2), year

(2009/2010). Plot **a** represents the values of each replicate and plot **b** summarizes data by the mean \pm SE in the two dimensions of the graph

The combination of the total amount of sugar and the ρ ratio values made it possible to discriminate between females that had fed before being caught in the field and females at the time of emergence (i.e. unfed) (Fig. 3). These two physiological parameters also made it possible to distinguish between *T. heterocerus* caught in the field at the beginning and at the end of flowering (Fig. 3).

Discussion

Floral resources in agroecosystems can stimulate natural enemy populations and the regulatory function they provide (Gurr et al. 2005; Heimpel and Jervis 2005). Indeed, it can increase natural enemies' efficiency by increasing longevity and fecundity, or changing the sex ratio (Berndt and Wratten 2005). However, knowledge about energy acquisition in the field of pollen beetle parasitoids had never been examined before.

The major finding of this study is that one of the most abundant and widespread univoltine parasitoids of the pollen beetle, *T. heterocerus*, feeds on sugar when foraging in the field. The overall content of sugar found in cocoons and foraging individuals suggests that (i) *T. heterocerus* emerge with relatively small amounts of sugar composed mainly of trehalose, glucose and fructose; (ii) the first parasitoids caught just after they appeared in the field had already consumed high amounts of sugar reserves; and (iii) the total amount of sugars then increases with the number of days since the beginning of oilseed rape flowering.

The presence of the disaccharide trehalose [the common insect haemolymph sugar, (Wäckers 2001)] in *T. heterocerus* at the time of emergence can be interpreted as the result of parasitoid larvae feeding on its host (i.e. the pollen beetle larva) just after pollen beetle pupation in the soil. Indeed, if it is assumed that *T. heterocerus* is a non host-feeding parasitoid at the adult stage, young parasitoid larvae developing in pollen beetle larvae have no lipid reserves in early development stages and, thus, feed on host haemolymph for maintenance and development (Jourdeuil 1960). In field-caught parasitoids the sugar spectrums never revealed trehalose. Steppuhn and Wäckers (2004) explained that absence of trehalose in adult parasitoids is surprising as it is reported in the literature that insects metabolize glucose into trehalose. Trehalose is assumed

to be involved in different functions such as energy storage, cryoprotection, thermal and osmotic stress, and feeding behaviour and nutrient intake (Thompson 2003). This difference in trehalose between just-emerged and field-caught individuals together with recent results on the nutritional state of other parasitoid species raises important questions about the role of trehalose and the success of functions usually assigned to trehalose (Steppuhn and Wäckers 2004; Vattala 2005). Further experiments on the role of trehalose involving different parasitoid species are therefore needed.

Flight is a highly energy-demanding behaviour and carbohydrates are known to constitute the main fuel for flight in most hymenopteran species (Casas et al. 2003; Wanner et al. 2006). Moreover, *T. heterocerus* emerges from previous year oilseed rape fields and is assumed to fly towards the host field by upwind anemotaxis, directly searching for hosts (Jourdeuil 1960; Williams et al. 2007). Hence, such flying patterns represent substantial energy consumption for parasitoids, which could explain why *T. heterocerus* individuals sampled at the beginning of flowering (i.e. just after arriving in the fields) have lower levels of sugar content than individuals at the time of emergence in 2009. Due to climatic conditions, the earliest sampling in 2010 occurred later than in 2009. Parasitoids may have been able to feed on sugar sources eight days after the beginning of flowering in 2010. This may explain why we found no difference in sugar content between specimens at emergence and at the beginning of flowering for 2010.

The combination of the total amount of sugar and the ρ ratio made it possible to discriminate between females that had fed before being caught in the field and females at the time of emergence (i.e. unfed), as the latter had significantly higher ρ ratios and significantly lower amounts of sugar than field-caught *T. heterocerus* females. However, the ρ ratios of males at the time of emergence showed high variability and it was not possible to differentiate between males caught in the field and males caught at the time of emergence. This is undoubtedly due to the fact that we found no sugars in one of the three pooled replicates for males at the time of emergence whereas the other two had relatively high ratios equivalent to the values of unfed females (ρ ratios = 1.00 and 0.65, Fig 3). As the HPAEC is able to detect very low amounts of sugars in the standards ($0.56 \mu\text{g ml}^{-1}$), we have strong confidence in the results displayed by our analyses.

All field-caught specimens that had fed (i.e. where we were able to quantify sugar amounts) had comparable ρ ratios (≈ 0.5), which were significantly lower than those of females in cocoons, and an overall content of sugar increasing with the number of days after the beginning of flowering. This indicates that *T. heterocerus* foraging in the field consume sugar and that adults caught at the end of flowering have consumed more sugar or have fed more recently than individuals at the beginning of flowering. These new results for this species are consistent with previous findings for other parasitoid species in which sugar consumption in the field have been found (Casas et al. 2003; Lavandero et al. 2005; Lee et al. 2006). Because the koinobiont parasitoid *T. heterocerus* does not feed on hosts during the imaginal stage and has never been observed feeding on honeydew (Jourdheuil 1960), our results strongly suggest that adults use nectar for carbohydrate acquisition after host field localization when foraging in the field and its surroundings.

Our results showed that at the end of flowering the sugar content of females was zero in 2009 and comparable to the sugar content of males at the beginning of flowering in 2010. These results are consistent with the knowledge about *T. heterocerus* biology and ecology. Jourdheuil (1960) reported that males always emerge a few weeks before females (protandry phenomenon, confirmed by our data in 2010) and that the sexes have very different behaviours. After emergence females immediately and actively forage for hosts in the field whereas males tend to fly in groups and can be found in the field and its immediate surroundings. Therefore, the higher amount of sugar found in males may be due to the fact that they have much more time for food-foraging in the crop and in adjacent habitats or that they consume less energy than the females actively seeking hosts.

Tersilochus heterocerus may use oilseed rape nectar as it represents an enormous amount of food in the landscape during flowering (Westphal et al. 2003). Nectars of numerous oilseed rape cultivars have been examined and provide unambiguous evidence that oilseed rape nectar is predominantly composed of glucose and fructose and that the total amounts of sugar are higher at the beginning of flowering than at the end (Pierre et al. 1999). However, *T. heterocerus* is present in oilseed rape fields until the end of flowering (Ulber et al. 2010; personal observation). Therefore, providing key floral resources within the field or in its

immediate vicinity towards the end of flowering might be a promising way to strengthen the top-down impact of parasitoid on the herbivore population.

This study provides the first evidence of the acquisition of carbohydrates in the field by *T. heterocerus*, one of the major parasitoid species of the pollen beetle. Several studies on various nectar-feeding species have demonstrated that providing floral resources increases longevity and fecundity of parasitoids and may enhance biological control of herbivores (Lee et al. 2006; Lee and Heimpel 2008; Bianchi and Wäckers 2008; Luo et al. 2010). We have provided evidence that *T. heterocerus* uses nectar when foraging in the field. Further research should explore the effect of various flowering plants (including oilseed rape) and sugars on longevity and fecundity of *T. heterocerus* and the subsequent parasitism rates of pollen beetle larvae. In a long-term perspective it can orientate breeders in selecting oilseed rape cultivars producing adapted amounts of sugars at key periods for parasitoids, such as the beginning of flowering, to strengthen biological control (Pierre et al. 1999; Tompkins et al. 2010). However, other aspects such as the effect of agronomic practices, characteristics linked to plant attractiveness, nectar accessibility, as well as trade-offs and synergies with other plant parameters should be clarified before including it in plant selection programs. Finally, studying the nutritional state of parasitoids in the field in different landscape contexts to examine the impact of floral and nectar resources in extra-field habitats should yield very interesting perspectives and major challenges for conservation biological control.

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